Preparation of chicken collagen-coated slides
An acid-soluble chicken collagen types-I and -III prep was prepared as described previously. Reconstitution of fibrillar chicken collagen was performed as described previously. Briefly, the collagen was dissolved in 0.5 M acetic acid at 1.5 mg/mL and dialyzed against 2 changes of 50 mM Tris HCl, pH 7.5 at 4°C over 48 h. After dialysis, the gelatinous white collagen precipitate was resuspended with a repeated aspiration through a 22-gauge needle and then sonicated with a steel probe sonicator (Branson 450 Digital Sonifer®, Danbury, CT) on ice for 3 pulses of 5 seconds at 10% amplitude with at least 15 seconds between pulses. Coating of microscope slides was performed as described previously. Briefly, 300 µL of fibrillar chicken collagen was placed in a flexiperm well that was attached to an EtOH-washed 25 × 75 mm glass (Fisher, Pittsburgh, PA) or Permanox™ (Nunc, Rochester, NY) microscope slide, for scanning electron microscopy assays, and incubated at 4°C overnight. Slides were then washed 2 times with PBS and blocked with 1% denatured BSA for at least 1 h at room temperature prior to use.

Real-time PCR analysis
Thrombocyte and lymphocyte RNA was isolated and cDNA was produced as described previously. SYBR Green master mix (Applied Biosystems, Carlsbad, CA) plus forward and reverse primers for chicken genes were used for real-time semiquantitative PCR, with each reaction performed in triplicate. PCR was performed on an Applied Biosystems 7900HT and results were analyzed with SDS 2.0 software. Data are expressed as ((Ct thrombocyte gene expression – Ct thrombocyte gapdh) – (Ct for lymphocyte gene expression – Ct lymphocyte gapdh)) where Ct is the cycle threshold and gapdh encodes glyceraldehyde dehydrogenase.
Primer pairs for the following chicken genes are listed below:

**gapdh**:  
Forward 5′ GAC AAC TTT GGC ATT GTG GAG GGT 3′  
Reverse 5′ AAG CTT CCC ATT CAG CTC AGG GAT 3′

**htr2a**:  
Forward 5′ TTG GGT ACC TTT CTT CAG CCG TCA 3′  
Reverse 5′ AGC TCT TCA TTT GAG CCA GGT GGA 3′

**mpl**:  
Forward 5′ CTC CTT CGA GGA TCT CAC GTG TTT 3′  
Reverse 5′ TGG TGG GTG CAT CCC AGT AGT AGA A 3′

**itgb3**:  
Forward 5′ TGC GGG AGG GAC TTC ATT GAG TTT 3′  
Reverse 5′ TGG ACA CGG AAC ATT TGG GAG TCA 3′

**gp9**:  
Forward 5′ AGC TTG CTC AGC CTG GAG GAA ATA 3′  
Reverse 5′ TGC TTC AAG GGC TTC ATC CTC ACA 3′

**f2r**:  
Forward 5′ ACA CGT GCC TCG GTG ATT TGT TTC 3′  
Reverse 5′ TGT GAA GCT CAG ATT CCC TCA GCA 3′

**selp**:  
Forward 5′ TGC TCT CTG GAA CGC TCA TTG TCT 3′  
Reverse 5′ AGT CGT AGG CTG CAT TGG TGA AGA 3′

**fv**:  
Forward 5′ TGC AGC TAA GGA AGT CTG CTG GAA 3′  
Reverse 5′ TGC TCT CTG GAA CGC TCA TTG TCT 3′

**p2ry12**:  
Forward 5′ TGA GAG GAT TTG TGT GCC AGG TCA 3′  
Reverse 5′ TGA ACG GTG AGG TGG CTT TCT TCT GAT 3′

REFERENCES

Figure S1. Thrombocyte adhesion to collagen under flow is integrin-dependent and sensitive to Src-family kinase inhibition

(A, B) Percent collagen surface area coverage of platelets (A) or thrombocytes (B) following perfusion of 5 mM EDTA-treated whole blood for 5 min was determined by analysis of fluorescent images. Shown are mean ± SEM, n = 3 experiments for each condition.
Figure S2
(A) Percent collagen surface area coverage of human platelets following perfusion of 10 µM eptifibatide-treated whole blood for 5 min was determined by analysis of fluorescent images. (B) Mean platelet aggregate area following perfusion of 10 µM eptifibatide-treated whole blood for 5 min. Shown are the mean ± SEM, n = 4 experiments for each condition.
Figure S3. Thrombus growth in the bird depends upon progressive FeCl₃ injury

The carotid arteries of anesthetized mice and Australian budgerigars (budgies) were exposed to filter pads soaked in the indicated concentrations of FeCl₃ for the indicated time periods. Transverse sections of injured carotid arteries were stained with hematoxylin-eosin (H–E) or Prussian blue (to detect FeCl₃). Note that platelet thrombi in the mouse carotid extend beyond the region of FeCl₃ injury while thrombi in the budgie carotid extend only as far as the depth of FeCl₃ injury. Scale bar = 50 µm. Shown are representative images, n = 1–5 experiments for each condition, as indicated by the number of diamonds and circles in Fig. 4A.